

Research Article

MORINDA MORINDOIDES (BACKER) MILNE-REDH (RUBIACEAE): PHYTOCHEMISTRY, PHARMACOLOGICAL **ACTIVITIES AND FUTURE DIRECTION: A MINI-REVIEW**

¹Sylvie Bolisomi Bongili, ^{2,*}Clément Liyongo Inkoto, ¹Néhémie Yamfu Bindanda, ¹Albert Bushabu Kongo, ²Blaise Mbembo wa Mbembo, ³Oscar IsekatsilaN'singa, ¹Andrien Masunda Tuwisana, ⁴Pius Tshimankinda Mpiana and ^{2,5}Koto-te-Nyiwa Ngbolua

¹Section of Laboratory Techniques, Higher Institute of Medical Techniques, Kinshasa, Democratic Republic of the Congo ²Department of Biology, Faculty of Sciences, University of Kinshasa, Kinshasa, Democratic Republic of the Congo ³Higher Pedagogical Institute of Boende, Tshuapa Province, Democratic Republic of the Congo ⁴Department of Chemistry, Faculty of Sciences, University of Kinshasa, Kinshasa, Democratic Republic of the Congo ⁵Department of Basic Sciences, Faculty of Medicine, University of Gbado-Lite, Democratic Republic of the Congo

Received 20th October 2020; Accepted 25th November 2020; Published online 30th December 2020

Abstract

The aim of this mini-literature review was to describe the traditional use, phytochemistry and biological properties of *M. morindoides*, a plant used as a conventional African medicine. In this investigation, the literature revealed that M. morindoides extracts were very efficient in the treatment of several diseases including malaria. According to our research, no studies have been conducted on the antisickling properties of Morinda morindoides. Further research is needed to assessment Antisickling activity of the different extracts of the parts of Morinda morindoides.

Keywords: Medicinal plant, Morinda morindoides, Traditional uses, Phytochemistry, Pharmacological activities.

INTRODUCTION

Nowadays, herbal medicine is an accepted and recognized form of medicine in the whole world (Mpiana et al., 2020). The plant produces various secondary metabolites which are bio-synthetically derived from primary metabolites, and these compounds are the main source of herbal pharmaceutical products (Tshilanda et al., 2019; Bongo et al., 2019; Panzu et al., 2020). The use of medicinal plants in therapy is a common practice in various parts of the world particularly in developing countries. According to the World Health Organization, a huge number of populations presently use phytomedicine for development of healthcare (Mohammed et al., 2020).60% of the global population is using the traditional medicine system in order to overcome several health related issue (Mohammed et al., 2020) and around 80% of African population use traditional medicine (Inkoto et al., 2020a; Inkoto et al., 2020b; Nkasa et al., 2020). Morinda morindoides (Rubiaceae) has been greatly appreciated in recent times and researchers have found scientific support for its use in folk medicine. Traditionally, the parts of this plant species have various applications, ranging from treatment of malaria, diarrhea, amoebiasis, constipation, intestinal parasites, rheumatic pains and fungus (Cimanga et al., 2006; Meite et al., 2009; Zirihi et al., 2010). Its parts of M. morindoides are reported to possess anti-inflammatory, cardiovascular. antimicrobial. antihypoglycemic, antihypertensive, anti-malaria, antispermatogenic, larvicidal, toxicological activities (Cimanga et al., 2006; Adenubi et al., 2010; Abdoulaye et al., 2017). In this article, we present the data on Morinda morindoides and its major chemical compounds that could justify their use in the treatment of Sickle Cell Anemia in the Democratic Republic of Congo. In addition, the objective of the current study is to review the literature on phytochemistry

and pharmacological activities of Morinda morindoidesused in Congolese traditional medicine. This data would allow the use of this plant as a multifunctional and low toxicity drug candidate for the management of various diseases, including the Sickle Cell Anemia.

METHODS

Search strategy

A non-exhaustive literature searches for relevant articles published in the last ten years was conducted in November 2020 on various electronic databases: Science Direct, Pubmed, Web of Science, Scopus, Google Scholar, POPLINE and System for Information on gray Litterature in Europe. As search strategy, the scientific name of this plant was used as a keyword, with the terms phytochemistry and Pharmacology. The chemical structures of *M. morindoides* naturally occurring compounds were drawn using ChemBioDraw Ultra 12.0 software package.

Eligibility criteria

In this literature mini-review, the selection of included studies was limited to studies with the inclusion criteria of:

- Phytochemistry data of M. morindoides, (i)
- Ethno pharmacological uses of this plant species in the (ii) past and clinical application.
- Pharmacological properties of M. morindoides in (iii) human, in vitro assay, or in vivo model. In our research, the studies obtained were excluded based on several criteria such as unreliably extracted data, only-abstract available.
- (iv) Overlapped data sets, reviews, thesis, editorials and book chapters.

^{*}Corresponding Author: Clément InkotoLiyongo, Department of Biology, Faculty of Sciences, University of Kinshasa, Kinshasa, Democratic Republic of the Congo.

RESULTS

Botanical description

Bagre et *al.*(2011) described *Morinda morindoides* (Fig.1) as a climbing liana, hairless with opposite leaves, elliptical oblong or elliptical obovate, wedge-shaped at the base. The leaves are 6-15 cm long and 3-8 cm wide. The leaf blade has about six pairs of lateral veins. The flowers, white in color, are grouped at the top, with the tube of the corolla short and robust. The fruits, bumpy, yellow when ripe, measure 4 cm in diameter.



Figure 1. Flowers and developing fruits of M. morindoides

Origin and geographic distribution

Morinda morindoides is distributed in a wide range of countries in West and Central Africa. It is native to Benin, Cabinda, Cameroon, Democratic Republic of Congo, Côte d'Ivoire, Nigeria, Liberia, etc. where it is called by several vernacular names such as Kongo bololo or Nkongabululu, Jologbo, ponju owiwi", zeleke, and brimstone (Akinloye *et al.*, 2015; Davis and Figueiredo, 2007).

Ethno-botanical uses

In southern African countries the parts of *Morinda morindoides* used to treat many diseases. The decoction of its leaves is traditionally used to treat certain parasitic diseases such as diarrhoea, amoebiasis, constipation and malaria (Meite *et al.*, 2009; Zirihi *et al.*, 2010). In addition, several studies have reported therapeutic uses of this plant such as antiplasmodial, antispermatogenic, antifungal (Kabangu, 1990; Williamson 2001; Bagre *et al.*, 2006; Mbatchi *et al.*, 2006; Adenubi *et al.*, 2010; Koffi *et al.*, 2010).

Phytochemistry

Several compounds have been isolated and/or identified in this plant by several authors including sterols, triterpenoids, glycosides, flavonoids and iridoids(Fig.2). Cimanga et *al.*(1995 and 1999) isolated flavonoid aglycones and glycosides from ethyl acetate and n-butanol extracts from the leaves of *Morinda morindoides*. These include quercetin, quercetin-7, 4'-dimethyl ether, quercetin-3-rhamnoside, quercetin-3-rutinoside (4), kaempferol-3-rhamnoside, kaempferol-3-rutinoside, Kaempferol-7-rhamnosysophoroside, luteolin-7-glucoside, apigenin-7-glucoside and chrysoerol-7neohesperidoside, kaempferol, apigenin, luteolin and chrysoerol. Eight iridoids were also isolated from ethyl acetate and n-butanol extracts from the leaves of this plant: gaiteroside, gaiteric acid, methoxygaiteroside, acetvl gaiteroside, epoxygaiteroside, epoxy methoxy gaiteroside, epoxy methoxy gaiteroside, dehydrogaiteroside, dehydromethoxygaiteroside (Cimanga et al., 2003;2006). Cimnga et al. (2008) isolated two antraquinones, notably alizarin and chrysarin, from the chloroformic extract of the leaves of this plant. Leaves of *M. morondoides* extracted and isolated a ketosteroid from petroleum ether: [(22E)-2ßhydroxy- 24- ethylcholesta- 4, 2,2- dien-3- one-1] (Harisolo etal. 2009). The work of Kouamé et al. (2010) has made it possible to highlight and identify, from the essential oils of this same plant species, fifty terpene compounds.

Biological activities

A. Antimalarial activity

Several antimalarial activities from Morinda morindoides were reported previously. Tona and Mesia (2001) evaluated in vivo antimalarial activity of ethanol, dichloromethane and lyophilized aqueous extracts of three plants, Cassia occidentalis (root bark), Morinda morindoides (leaves) and Phyllanthus niruri (whole plants) against Plasmodium berghei ANKA in mice. The result of this study indicated that dichloromethane extracts of this species have an antimalarial activity. Each lyophilized aqueous extract was less active than the corresponding ethanolic extract. Tona et al. (2004) reported in vitro antiplasmodial activity of seven ethanol extracts and twenty fractions of the partitioning of the initial ethanol extracts of seven African medicinal plants used in Democratic Republic of Congo (DRC) in treatment of malaria. The ethanol extracts from the leaves of Morinda morindoides (50 < IC50 <100g/mL) were less active, but their petroleum ether fraction showed pronounced antiplasmodial activity (IC₅₀ < 3g/mL). According to the authors, the observed antiplasmodial activity could be related to the presence of terpenes, steroids, coumarins, flavonoids, phenolic acids, lignans, xanthones and anthraquinones. Antidiarrheal activity. Meite et al. (2009) studied the effects of ethyl acetate extract of Morinda morindoides against castor oil-induced experimental diarrhea in albino Wistar rats. The results of this study indicated that at oral doses of 250, 500 and 1000 mg/kg body weight, the plant extract showed pronounced and dose-dependent antidiarrheal activity. The protective role of the extract at 1000 mg/kg was comparable to that of the reference drug, loperamide (5mg/kg). The extract (1000 mg/kg) produced a decrease in intestinal transit similar to that of atropine (5mg/kg), and significantly (p<0.01) inhibited castor oil-induced enteropathy.

B. Antiamoebic effect

Cimanga *et al.*(2006) evaluated the potential anti-amoebic activity and cytotoxic effect of aqueous decoction, 10 flavonoids and 4 iridoids isolated from 80% methanol extract of *M. moridoides* leaves against MT-4 cells. The results indicated that the aqueous decoction and the 80% Methanol extract showed interesting anti-amoebic activity with IC₅₀ values of 3.1 ± 1.7 and 1.7 ± 0.6 g/mL respectively. Apigenin-7-O-glucoside and luteolin-7-O-glucoside showed moderate anti-amoebic activity with IC₅₀ values of 22.3 ± 3.2 and 37.4 ± 2.7 g/mL, respectively. Kaempferol(IC₅₀=10.3±2.3g/mL), apigenin (IC₅₀=12.7±4.3g/mL) and luteolin (IC₅₀ = 17.8±4.3g/mL) showed more pronounced activity than their corresponding glycosides. All iridoids tested showed very good activity with IC₅₀ values below 10g/mL.



Figure 2. Cheminical structures of Morindamorindoides

The most active iridoids were epoxygaertneroside (IC₅₀ = 1.3 ± 0.4 g/mL) and methoxygaertneroside (IC₅₀ = 2.3 ± 0.7), followed by gaertneroside and gaerneric acid with IC₅₀ values of 4.3 ± 1.8 and 7.1 ± 1.4 g/mL, respectively. Except for quercetin and 7,4-dimethyl quercetin ether which showed a cytotoxic effect with IC₅₀ values ranging from 14 to 22 g/mL.In another study, Tona et *al.* [1998], in which several plants from the Democratic Republic of Congo were tested *in vitro* for their Antiamoebic activity, the aqueous extract from the leaves of *Morinda morindoides* showed a very high inhibitory activity (MIC=15.6 µg/mL).

C. Antibacterial activity

Antibacterial activity was evaluated used a soap containing hexane extract from Morinda morindoidesleaves (Abdoulaye et al., 2017). The results of this study showed that has the soap containing hexane extract of this plant have an antibacterial effect. The base soap had the lowest bactericidal effect with a MIC of 62.50 mg/mL for the two strains tested and MBC of 125.00 mg/mL and 62.50 mg/mL for Staphylococcus aureus and Pseudomona aeruginosa respectively. The results led to the identification of compounds all belonging to the anthraquinone family. The antibacterial activity of anthraquinone was tested against six strains of microorganisms using ofloxacin as standard (Moroh et al., 2019). The minimum inhibitory concentrations+) recorded ranged from 8 to 128 µg/mL and Staphylococcus aureus was the most sensitive organism. Moroh et al. (2008) evaluated in vitro the antibacterial activity of the acetatic extract of fresh leaves of M. morindoides (Baker) Milne Redheat (Rubiaceae) on eight (8) strains of Escherichia coli, which are commonly found in diarrhea in children aged 0 to 5 years. Of these eight strains, three are reference strains namely Escherichia coli ATCC 25922, Escherichia coli ATCC 8739 and Escherichia coli 2361; three of known serotype (Escherichia coli O26H6, Escherichia coli O142 K86 and Escherichia coli O126 B16); isolated from well water and one of hospital origin. The extract was effective on all these strains. The Minimum Inhibitory Concentrations (MIC) recorded ranged from 3.75 to 15 mg/mL while the Minimum Bactericidal Concentrations (MBC) were found to be between 7.5 and 30 mg/mL. Considering this positive result, the authors recommend the use of M. morindoides leaves extract as a treatment for children's diarrhea.

D. Anti-spermatogenic activity

Adenubi et *al.* (2010) reported the antispermatogenic activity of aqueous extract of *Morinda morindoides* root bark on six male wistar rats. *M. morindoides* root bark extract caused a significant (p<0.05) reduction in sperm motility, a significant dose-dependent reduction in sperm count and a significant (p<0.05) dose-dependent increase in morphological abnormalities of sperm in treated rats. Thus, this study concludes that root bark extract of this plant has significant antispermatogenic effects on adult male Wistar rats, which may affect the reproduction of these male Wistar rats.

E. Cardiovascular activity

Gboko *et al.* (2012) investigated the properties of total protein extracts from the leaves of *Morinda morindoides* for rabbit blood pressure and mechanical activity of isolated rat hearts. The results showed that the total protein extract of M.

morindoides has a blood pressure reducing effect $(31.77 \pm 2.72\%)$ that would result from the combined actions of myocardial depression and muscarinic cholinergic receptor activation mediated by vascular smooth muscle relaxation.

F. Hypoglycaemic and hypolipidaemic activities

Olukunle et al.(2012) tested the hypoglycemic and hypolipidemic effects of M. morindoides root bark extracts and fractions. The aqueous and methanolic extracts were administered orally to 48 rats at a dose of 400 mg kg for 21 days. Fractions (hydromethanol, h hexane, chloroform and ethyl acetate) from bioactivity guided fractionation and A-F (CsF) subfractions of accelerated gradient chromatography were also evaluated in 45 rats for hypoglycemic activity at doses of 400 mg/kg, 200 mg/kg and 100 mg/kg respectively. Glibenclamide was used as a positive control. Polyoxyethylene sorbitan monooleate and distilled water administered to rats were used as negative controls. Morinda morindoides 400 mg/kg of aqueous and methanolic extracts and 100 mg/kg of chloroform CsF B caused (62.8%, 56% and 74%, respectively) reductions in blood glucose (BGL) levels. The aqueous extract caused a significant (P < 0.05) decrease in serum cholesterol values (133.48 ± 1.10) mg-dl', low density lipoproteins (66.38 \pm 2.50) mg/dL and a significant (P < 0.05) increase in high density lipoproteins (51.0 ± 3.0) mg/dL compared to control.

G. Antifungal activity

Bagre et al. (2006, 2011) reported that ethyl acetate extract of M. morindoides leaves possess antifungal effects against Cryptococcus neofoman with IC₅₀ of 1.35 mg/mL. Touré et al.(2010) developed a soap formulation based on hexane extract from the leaves of Morinda morindoides in order to evaluate its antifungal activity against human-derived fungalisolates. The result showed that M. morindoides extract soap inhibited the growth of all fungal strains at a MIC of 31.25 mg/mL. In addition, the base soap (control) also Candida albicans at a MIC of 125 mg/mL and at a MIC of 62.50 mg/mL for the other 3 strains tested. Touré et al. (2011) compared the antifungal activity of various extracts (aqueous, ethanol, ethyl acetate and hexane) of M. morindoides and performed phytochemical screening. The results showed a higher antifungal activity of the hexane extract against C. albicans with a MIC of 31.25 mg/mL and IC₅₀ of 6.17±1.04 mg/mL compared to the other extracts (aqueous, ethanolic and acetate). The same extract showed the highest activity against T. rubrum with a MIC of 15.62 mg/mL and an IC₅₀ of 2.68±1.19 mg/mL.

H. Cytotoxicity

Regarding cytotoxicity effect, Marie-Genevieve et *al.* (2010) claimed that toluene extract of *M. morindoides* leaves dispayed a potent effect on P388 and L1210 cells with IC_{50} =6.0 and 6.5 µg/mL, respectively, while K562 cells were slightly less sensitive to the exposure of toluene extract 12.2 µg/mL. Nevertheless, extraction with methyl tert-butyl ether (MtBE) was less efficient than toluene extract in all tested cell lines. The effects of toluene and MtBE extracts in K562 human erythroleukemia cells may be due to apoptotic induction. On MT4 cells, Cimanga *et al.* (2006) reported that quercetin and quarcetin-7, 4'-dimethylether produced cytotoxic effect with CC_{50} ranging from 14 to 22μ g/mL. Interestingly, *M. morindoides* appeared not to damage normal cells. At high

concentration of 150 μ M, some phenylpropanoid conjugated iridoids caused 13.4% cytotoxic in maximum (Zirihi *et al.*, 2005). Boga *et al.* (2105) indicated that, dichloromethaneethanol extract of this plant, showed relativy low toxicity in normal rats with LC₅₀ within 200-5000 mg/kg. Moreover, with doses ranging 50-100 mg/kg, this extract exerted no significant effect on body weight, biochemical parameters and blood parameters. Unfortunately, these studies used different extracts from *M. morindoides*, in which constituents were not specified so that we could not conclude whether *M. morindoides* only caused toxicity on abnormal cell lines.

Conclusion

In the current mini-review the aim was to review the literature on the traditional use, phytochemistry and biological properties of this valuable plant species. The results of this bibliographic investigation revealed that *M. morindoides* contains pharmacologically active substances as sterols, triterpenoids, glycosides, flavonoids and iridoidspharmacologically active substances with antidiarrheal, antimalarial, Antispermatogenic, antiamoebic properties. According to our research, no studies have been conducted on the antisickling properties of *Morinda morindoides*. Further research is needed to assessment Antisickling activity of the different extracts of the parts of *Morinda morindoides*.

REFERENCES

- Abdoulaye, T., Kouakou, S.K., Bakary, C., Karamoko, O. andCoulibaly, A. 2017. Antibacterial Activity of Soap containing Hexane Extract of Leaves of Morinda morindoides (Morinda: Rubiaceae) against Staphylococcus aureusand Pseudomonas aeruginosa, *Int.J.Curr.Microbiol.App.Sci.*, 6(1): 512-517.
- Adenubi, O., Olukunle, J., Abatan, M., Ajayi, O., Adeleye, O. and Kehinde, O. 2010. Antispermatogenic activity of morinda morindoides root bark extract in male wistar rats, *Journal of Natural Sciences Engineering and Technology*, 9: 99-105.
- Adenubi, O.T., Olukunle, J.O., Abatan, M.O., Ajayi, O.L., Adeleye, O.E. and Kehinde, O.O. 2010. Antispermatogenic activity of morinda morindoides root bark extract in male wistar rats, J. Nat. Sci. Engr. Tech., 9(1):99-105.
- Akinloye, D.I., Sunmonu, T.O., Omotainse, S.O. and Balogun, E.A. 2015. Evaluation of antioxidant potentials of Morinda morindoides leaf extract, *Toxicological & Environmental Chemistry*, 97(2): 155–169.
- Atindehou, K.K., Schmid, C., Brun, R., Koné, M.W. and Traore, D. 2004. Antitrypanosomal and antiplasmodial activity of medicinal plants from Cote d'Ivoire, *Journal of Ethnopharmacology*,90:221–227
- Bagre, I., Bahi, C., Meite, S., Djaman, A. and Guede, G. 2006. Evaluation et amelioration in vitro de l'activité antifongique de Morinda morindoides (Baker) milneredh (Rubiaceae) sur Cryptococcus noeformans, un champignon responsable de mycose humaine. *Journal of Scientific*, 7:37-46.
- Bagre, I., Bahi, C., Ouattara, K., Denotes, N., Djaman, A., Coulibaly, A. and N'Guessan, J. 2011. Etude botanique et exploration de l'activité antifongique de morinda morindoides (Baker) milne-redh. sur la croissance in vitro de cryptococcus neoformans, Phytotherapie, 9: 136-141.
- Boga, G., Bahi, C., Konkon, N., Yapi, H., Djama, A. and Niguessan, J. 2015. Assessment of acute and subacute

toxicity of the total dichloromethane-ethanol extracts of morinda morindoides (Baker) milne-redh on rats. Pharmacognosy Magazine 7 (6).

- Bongo, G., Inkoto, C., Masengo, C., Tshiama, C., Lengbiye, E., Djolu, R., Mutwale, K., Ngombe, K., Mbemba, T., Tshilanda, D., Mpiana, P. and Ngbolua, K.N. 2017.
 Antisickling, Antioxidant and Antibacterial Activities of Afromomum alboviolaceum (Ridley) K. Schum, Annona senegalensis Pers. and Mondia whitei (Hook. f.) Skeels, American Journal of Laboratory Medecine, 2(4): 52-59.
- Cimanga, K., De Bruyne, T., Hu, J.P., Cos, P., Apers, S., Pieters, L., Tona, L., Kambu, K., Vanden Berghe, D. and Vlietinck, A.J. 1999.Constituents from *Morinda morindoides* leaves as inhibitors of xanthine oxidase and scavengers of superoxide anions, *Pharmacy and Pharmaco/ogy Communications;* 5(6): 419-424.
- Cimanga, K., De Bruyne, T., Lasure, A., Li, Q., Pieters, L., Claeys, M., Vanden Berghe, D., Kambu, K., Tona, L. and Vlietinck, A.1995.Flavonoid 0-glycosides from the leaves of *Morinda morindoides, Phytochemistry*; 38(5): 1301-1303.
- Cimanga, K., Hermans, N., Apers, S., Van Miert, S., Vanden Heuvel, H., Claeys, M., Pieters, L., Vlietinck, A.J.2003.Complement-inhibiting iridoids.fi"om Morinda morindoides, Journal of Natural Products; 66, 97-102.
- Cimanga, K.R., Kambu, K., Tona, L., Hermans, N., Apers, S., Totté, J., Pieters, L. and Vlietinck, A.J.2006.Cytotoxicity and *in vitro* susceptibility of *Entamoeba histolytica* to *Morinda morindoides* Jeaf extracts and its isolated constituents, *Journal of Ethnopharmacology*, 107 : 83-90.
- Cimanga, K.R., Tona, L.G., Kambu,K.O., Mesia,K.G., Muyembe,T.J.J.,Apers,S., Pieters,L., Vlietinck,A.J. 2008. Antimalarial activity of some extracts and isolated constituent from leaves *Morinda morindoides*. *Journal of Natural Remedies* 8/2, 191-202.
- Cimanga, R.K., Kambu, K., Tona, L., Hermans, N., Apers, S., Totté, J., Pieters, L., Vlietinck, A.J. 2006. Cytotoxicity and in vitro susceptibility of Entamoeba histolytica to Morinda morindoides leaf extracts and its isolated constituents, *Journal of Ethnopharmacology*, 107(1):83-90
- Figueiredo, E. 2007. The Rubiaceae of Cabinda (Angola). Botanical Journal of the Linnean Society, 154(4):455–495.
- Gboko, A.O., M'boh, G.M., Coulibaly, F.A., N'Guessan, J.D., Djaman, A.J., Coulibaly, A. 2012. Cardiovascular activity of total protein extract of Morinda morindoïdes (baker) Miln-Redhon on rabbit blood pressure and isolated rat heart. *Afr. J. Pharm. Pharmacol.*, 6(12):871-876.
- Harisolo, R., Seri, C.S., Yao, K.P.A., Okpekon, T., Adiko, V.A., Djakoure, L.A., Coffy, A.A. 2009.A Ketosteroid Isolated from *Morinda Morindoides, European Journal of Scientific Research*,28 (4): 621-626.
- Inkoto, C.L., Kayembe, J.P.K., Mpiana, P.T., Ngbolua, K.N. 2020a. A review on the Phytochemistry and Pharmacological properties of Picralima nitida Durand and H. (Apocynaceae family): A potential antiCovid-19 medicinal plant species, *Emer Life Sci Res.*, 6(1): 64-75
- Inkoto, C.L., Masengo, C.A., Falanga, C.M., Membo, W.M.B., Amogu, J.J.D., Mahendra, I.S.M., Kayembe, J.P.K., Mpiana, P.T., Ngbolua, K.N. 2020b. A Mini-Review on the Phytochemistry and Pharmacology of the Plant Carica Papaya L. (Caricaceae). *Britain International of Exact Sciences (BIoEx)*, 2(3): 663-675.
- Kabangu, K. 1990. Eléments de phytotherapie comparée : plantes médicinales africaines. Centre de recherches pédagogiques

- Koffi, A., Yapi, H., Bahi, C., Guessend, K., Djaman, J., Guede-Guina, F.2010. Antimicrobial activity of morinda morindoides on in vitro growth of vibrio cholerae in cote d'Ivoire. *Med Trop (Mars)*, 70, 53-56.
- Kouame, B.F.P., Bedi, G., Koffi, A.M., Chalchat, J.C. et N'guean, T.Y. 2010. Volatiles Constituents from Leaves of *Morinda morindoides* (Rubiaceae): A Medicinal Plant from the Ivory Coast, *The Open Natural Products Journal*,3: 6-9.
- Marie-Genevieve, O., Robin, O., Gregory, G., Attibayeba, Catherine, L., Claudie, M.2010. Cytotoxic effect induced by morinda morindoides leaf extracts in human and murine leukemia cells, African Journal of Biotechnology; 9: 6560–6565.
- Mbatchi, S., Mbatchi, B., Banzouzi, J., Bansimba, T., Nsonde, N.G. and Ouamba, J.2006. In vitro antiplasmodial activity of 18 plants used in congo brazzaville traditional medicine, *Journal of Ethnopharmacology*, 104 : 168–174.
- Meite, S., N'guessan, J.D., Bahi, C., Yapi, H.F., Djaman, A.J., Guede, G.V. 2009. Antidiarrheal activity of the Ethyl Acetate Extract of *Morinda morindoides* in Rats. *Trop J Pharm Res*, 8 (3): 201-207.
- Moroh, J.L.A., Bahi, C., Dje, K., Loukou, Y.G., Guede-Guina, F. 2008. Study of the antibacterial activity of Morinda morindoides (Baker) milne-redheat (rubiaceae) acetatique extract (ACE) on in-vitro growth of Escherichia coli strains, Bulletin de la Société Royale des Sciences de Liège, 77 (2) :44- 61.
- Moroh, J.L.A., Fleury, Y., Coulibaly, A., Labia, R., Leguérinel, I. 2019. Chemo-Diversity of Antibacterial Anthraquinones from the Roots of Morinda morindoides. *The Natural Products Journal*, 9(4): 256-261.
- Mpiana, P.T., Ngbolua, K.N., Tshibangu, D.S.T., Kilembe, J.T., Gbolo, B.Z., Mwanangombo, D.T., Clement, L. Inkoto, Lengbiye, E.M., Mbadiko, C.M., Matondo, A., Bongo, G.N. and Dorothée D. Tshilanda, 2020. Aloe vera (L.) Burm. F. as a Potential Anti-COVID-19 Plant: A Minireview of Its Antiviral Activity, *EJMP*, 31(8): 86-93
- Nkasa, H.L., Clément Inkoto, C.L., Muzomwe, M.J.C., Masengo, E.I., Mumba, C.M., Mulenga, C.M. and Taba, K.M. 2020. Phytochemical Screening and Antibacterial Activity of Phytomedecine Mathesia, A Drug use against Buruli Ulcer in Republic Democratic of the Congo (Drc), Ejpmr; 7(2), 52-56
- Olukunle, J.O., Abatan, M.O., Adenubi, O.T., Amusan, T.A. 2012. Hypoglycaemic and hypolipidaemic effects of crude extracts and Chromatographie fractions of Morinda morindoides root bark in diabetic rats, *Acta Vet. Brno.*, 81:269-274

- Panzu, N.Z., Inkoto, C.L., Ngbolua, K.N., Mukeba, F.B., Kitadi, J.M., Taba, K., Mbala, B.M., Tshilanda, D.D., Kayembe, J.P.K. and Pius Mpiana, T. 2020. Review on the phytochemistry, toxicology and bioactivities of *Euphorbia hirta* L.: A potential antisickling medicinal plant species, JMPHTR, 7: 8-18
- Tona, L., Cimanga, R.K., Mesia, K., Musuamba, C.T., De Bruyne, T., Apers, S., Hernans, N., Van Miert, S., Pieters L, Totté, J., Vlietinck, A.J. 2004. In vitro antiplasmodial activity of extracts and fractions from seven medicinal plants used in the Democratic Republic of Congo, *Journal* of Ethnopharmacology, 93(1):27-32.
- Tona, L., Kambu, K., Ngimbi, N., Cimanga, K., Vlietinck, A.J. 1998. Antiamoebic and phytochemical screening of some Congolese medicinal plants, *Journal of Ethnopharmacology*, 61:57-65
- Tona, L., Mesia, K. 2001. In-vivo antimalarial activity of Cassia occidentalis, Morinda morindoides and Phyllanthus niruri. Annals of Tropical Medicine & Parasitology, 95(1):47-57
- Toure, A., Bahi, C., Bagre, I., N'Guessan, J., Djaman, A., Coulibaly, A. 2010. In vitro antifungal activity of the soap formulation of the hexane leaf extract of morinda morindoides (Morinda; Rubiaceae), *Tropical Journal of Pharmaceutical Research*, 9(3):237-241.
- Touré, A., Bahi, C., Ouattara, K., Djama, A.J., Coulibaly, A. 2011. Phytochemical screening and in vitro antifungal activities of extracts of leaves of Morinda morindoides (Morinda, Rubiaceae), J. Med. Plants Res., 5(31):6780-6786.
- Toure, A., Bahi, C., Ouattara, K., Djama, J., Coulibaly, A. 2011. Phytochemical screening and in vitro antifungal activities of extracts of leaves of morinda morindoides (Morinda, Rubiaceae), *Journal of Medicinal Plants Research*, 5 : 6780-6786.
- Tshilanda, D.D., Inkoto, C.L., Kashala, M., Mata, S., Mutwale, P.K., Tshibangu, D.S.T., Bongo, G.N., Ngbolua, K.N., Mpiana, P.T. 2019. Microscopic Studies, Phytochemical and Biological Screenings of Ocimum canum; International Journal of Pharmacy and Chemistry, 5(5): 61-67
- Williamson, E. 2001. African Traditional Medicine-A Dictionary of Plant Use and Applications. Medpharm Scientific Publishers.
- Zirihi, G., N'guessan, K., Etien, D., Grellier, P. 2010. Ethanopharmocological study of plant used to treat malaria, in traitional medicine, by bete populations of ISSIA (Cote d'Ivoire), *International Journal of Pharmaceutical Sciences* and Research, 2(4): 216–217.